



Hot Start *Taq* DNA Polymerase

Catalog #	3291	3293
Package Size	200 units	1,000 units
Volume	40 µl	200 µl
Concentration	5 units/µl	

Catalog #	3291d	3293d
Package Size	200 units	1,000 units
Volume	40 µl	200 µl
Concentration	5 units/µl	

*Catalog numbers ending with "d" include separate dNTP mix.

Description

Intact Genomics Hot start *Taq* is a thermostable DNA polymerase that possesses a 5' → 3' polymerase activity (1, 2) and a 5' flap endonuclease activity (3, 4). Hot Start *Taq* DNA Polymerase is chemically modified that leads to complete inactivation of the polymerase until the initial heat activation step at the start of PCR. Hot start PCR reduces non-specific amplification during setup stages of the reaction and helps increase PCR specificity and sensitivity. This product is supplied with the unique Intact Genomics 10x PCR reaction buffer, containing MgCl₂, which produces a final Mg²⁺ concentration of 1.5 mM, and 5X Magic Enhancer that enables efficient amplification of GC rich templates up to 84%.

Protein Purity

The physical purity of this enzyme is ≥98% as assessed by SDS-PAGE with Coomassie® blue staining (see figure below).

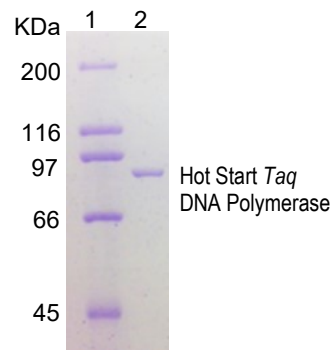


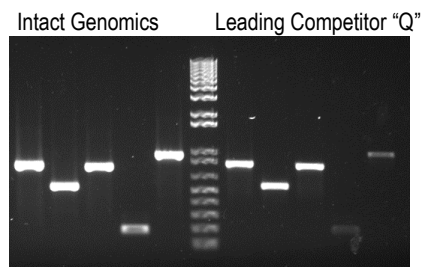
Fig. 1: Lane 1. Protein Marker
Lane 2. Hot Start *Taq* DNA Polymerase

Product Source

E. coli strain expressing a *Taq* DNA Polymerase gene from *Thermus aquaticus* YT-1.

Hot start *Taq* Polymerase Comparison Data:

We compare our Hot Start *Taq* Polymerase with leading competitor side by side and our enzyme is better than the competitor. A typical gel picture is shown below:



Applications

- Routine PCR
- Primer extension
- Colony PCR
- Efficient for amplifying high GC template DNA with Magic Enhancer

Product Includes

- 1) Hot Start *Taq* DNA Polymerase
- 2) 10x PCR Buffer with Mg²⁺
- 3) 5x Magic Enhancer
- 4) 10 mM dNTP (Cat. # 3291d, 3292d only)

Storage Temperature

-20 °C

Storage Buffer

50 mM Tris-HCl, 50 mM KCl, 1 mM DTT, 0.1 mM EDTA, 50% Glycerol, pH 7.5 @ 25 °C

10x PCR Buffer with Mg²⁺

100 mM Tris-HCl pH 9.0, 15 mM MgCl₂, 100 mM KCl, 80 mM (NH₄)₂SO₄, 0.5% Igepal CA 630

Unit Definition

One unit is defined as the amount of enzyme that incorporates 10 nmoles of dNTP into acid-insoluble form in 30 minutes at 72 °C.

Protocol

1. Thaw 10x PCR Buffer, dNTP mix, primer solutions, 5X Magic Enhancer (if required) and mix thoroughly before use.
2. Prepare a reaction mix according to the following table:
The reaction mix typically contains all the components needed for PCR except the template DNA.

PCR Reaction Set Up:	
Template DNA	x µl (0.01-0.5 µg)
10x PCR Buffer	10.0 µl
dNTP (10 mM)	2.0 µl
Forward Primer	x µl (0.1- 0.5 µM)
Reverse Primer	x µl (0.1- 0.5 µM)
5x Magic Enhancer (optional)	20 µl
Hot start <i>Taq</i> DNA Polymerase (5 U)	0.5 µl
H ₂ O up to	100.0 µl

3. Mix the reaction mixture thoroughly.
4. Add template DNA to the individual PCR tubes containing the reaction mixture.
5. Program the thermal cycler according to the manufacturer's instructions.

Note: PCR program must start with an initial heat-activation step at 95 °C for 15 min.

A typical PCR cycling program is outlined in the following table.

PCR Cycling Conditions:			
Steps	Temp.	Time	Cycles
Initial denaturation	95 °C	15 min	1
Denaturation	94 °C	30-60 sec	25-35
Annealing	52-66 °C	30-60 sec	
Extension	72 °C	1-2 min	
Final extension	72 °C	10 min	1
Hold	4-12 °C	∞	

6. Place the PCR tubes in the thermal cycler and start the cycling program.

Reference

1. EChien, A., Edgar, D.B. and Trela, J.M. (1976). J. Bact. 127, 1550-1557.
2. Lawyer, F.C. et al. (1993). PCR Methods and Appl. 2, 275-287.
3. Longley, M.J., Bennett, S.E. and Mosbaugh D.W. (1990). Nucleic Acids Res. 18, 7317-7322.
4. Lyamichev, V., Brow, M.A. and Dahlberg, J.E. (1993). Science. 260, 778-783.