

ig™ *Lactococcus lactis* Cells Electrocompetent Cells

Description

Intact Genomics Electroporation Competent *Lactococcus lactis* Cells are customer strains optimized for the highest transformation efficiencies which is ideal for applications requiring high transformation efficiencies, such as with cDNA or gDNA library construction.

Specifications

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| Competent cell type: | Electrocompetent |
| Species: | <i>L. lactis</i> |
| Strain: | IL 1403 or MG1363 |
| Format: | Tubes |
| Transformation efficiency: | $\geq 1 \times 10^6$ cfu/ μ g pNZ8148 DNA |
| Blue/white screening: | No |
| Shipping condition: | Dry ice |

Reagents Needed for One Reaction

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| ElectroCompetent <i>Lactococcus lactis</i> Cells : | 25 μ l |
| DNA (or pNZ8148 Control, 50 ng/ μ l): | 1 μ l |
| Recovery medium: | 1 ml |

Storage

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| ElectroCompetent <i>Lactococcus lactis</i> Cells : | -80 °C |
| pNZ8148 control DNA: | -20 °C |
| Recovery medium: | 4 °C |

Quality Control

Transformation efficiency is tested by using the pNZ8148 control DNA supplied with the kit and using the protocol in this manual. Transformation efficiency should be $\geq 1 \times 10^6$ CFU/ μ g pNZ8148 DNA. Untransformed cells are tested for appropriate antibiotic sensitivity.

General Guidelines

Follow these guidelines when using Electroporation Competent *Lactococcus lactis* Cells:

- Handle competent cells gently as they are highly sensitive to changes in temperature or mechanical lysis caused by pipetting.
- Thaw competent cells on ice, and transform cells immediately following thawing. After adding DNA, mix by tapping the tube gently. Do not mix cells by pipetting or vortexing.

Note: A high-voltage electroporation apparatus such as Bio-Rad Gene Pulser II #165-2105, capable of generating field strengths of 16 kV/cm is required.

Calculation of Transformation Efficiency

Transformation Efficiency (TE) is defined as the number of colony forming units (cfu) produced by transforming 1 μ g of plasmid into a given volume of competent cells.

$$TE = \text{Colonies}/\mu\text{g}/\text{Dilution}$$

Transform 1 μ l of (50 ng/ μ l) pNZ8148 control plasmid into 25 μ l of cells, add 976 μ l of Recovery Medium. Dilute 100 μ l of this in 900 μ l of Recovery Medium and plate 50 μ l. Count the colonies on the plate in two days. If you count 300 colonies, the TE is calculated as follows:

$$\begin{aligned} \text{Colonies} &= 300 \\ \mu\text{g of DNA} &= 0.05 \\ \text{Dilution} &= 50/1000 \times 100/1000 = 0.005 \\ \text{TE} &= 300/.05/.005 = 1.2 \times 10^6 \end{aligned}$$

Transformation Protocol

Use this procedure to transform Electroporation Competent *Lactococcus lactis* Cells. Do not use these cells for chemically transformation.

- 1) Place sterile cuvettes and microcentrifuge tubes on ice.
- 2) Remove competent cells from the -80 °C freezer and thaw completely on wet ice (10-15 minutes).

- 3) Aliquot 1 μ l (1 ng -10 μ g) of DNA to the chilled microcentrifuge tubes on ice.
- 4) When the cells are thawed, add 25 μ l of cells to each DNA tube on ice and mix gently by tapping 4-5 times. For the pNZ8148 control, add 1 μ l of (50 ng/ μ l) DNA to the 25 μ l of cells on ice. Mix well by tapping. Do not pipette up and down or vortex to mix, this can harm cells and decrease transformation efficiency.
- 5) Pipette 26 μ l of the cell/DNA mixture into a chilled electroporation cuvette without introducing bubbles. Quickly flick the cuvette downward with your wrist to deposit the cells across the bottom of the well and then electroporate.
- 6) Immediately add 976 μ l of Recovery Medium or any other medium of choice to the cuvette, pipette up and down three times to re-suspend the cells. Transfer the cells and Recovery Medium to an Eppendorf tube.
- 7) Seal the closed tube caps with parafilm and quickly warm tubes to 37 °C using a water bath.
- 8) Incubate tubes at 37 °C for 3 hours with no shaking.
- 9) Dilute the cells as appropriate then spread 20-200 μ l cells onto a pre-warmed selective plate. For the pNZ8148 control, dilute the cells 1/10 and plate 50 μ l of diluted transformants onto an MRS plate containing 10 μ g/ml chloramphenicol. Use sterilized spreader or autoclaved ColiRoller™ plating beads to spread evenly.
- 10) Incubate the plates for 2 days at 37 °C under anaerobic conditions.

Electroporation settings

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| Mode | Exponential protocol |
| Voltage (V) | 2,500 V |
| Capacitance | 25 μ FD |
| Resistance | 400 ohms |
| Cuvette | 2 mm |